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[7]	L7	L4	30
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	L6	L4 and (ankara modified virus)	1
M	L5	L4 and avipox	1
!	L4	L3 and vaccinia	58
V	L3	L2 and vector	94
	L2	L1 and envelope protein E1	129
	L1	HCv	6460

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                 and text labels
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       3326456 "C"
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          9134 "ENVELOPES"
         56544 "ENVELOPE"
                  ("ENVELOPE" OR "ENVELOPES")
       1761714 "PROTEIN"
       1225322 "PROTEINS"
       2047023 "PROTEIN"
                 ("PROTEIN" OR "PROTEINS")
         35928 "E1"
             O "HEPATITITS C ENVELOPE PROTEIN E1"
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       3326456 "C"
        321999 "VIRUS"
         68677 "VIRUSES"
        333808 "VIRUS"
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         51183 "ENVELOPE"
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         56544 "ENVELOPE"
                  ("ENVELOPE" OR "ENVELOPES")
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=> HCV
          9348 HCV
            18 HCVS
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          9134 ENVELOPES
         56544 ENVELOPE
                 (ENVELOPE OR ENVELOPES)
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           674 L3 AND L4
=> recombinant and L5
        173012 RECOMBINANT
          6652 RECOMBINANTS
        176631 RECOMBINANT
                 (RECOMBINANT OR RECOMBINANTS)
           154 RECOMBINANT AND L5
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        89478 VECTORS
        198198 VECTOR
                 (VECTOR OR VECTORS)
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           288 RVV
            42 RVVS
           300 RVV
                 (RVV OR RVVS)
Г8
             0 RVV AND L7
=> vaccinia and L7
          9963 VACCINIA
             2 VACCINIAS
          9964 VACCINIA
                 (VACCINIA OR VACCINIAS)
L9
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=> truncated and L8
         32137 TRUNCATED
L10
             O TRUNCATED AND L8
=> truncated and L7'
         32137 TRUNCATED
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             5 TRUNCATED AND L7
=> glycosylation and L7
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           503 GLYCOSYLATIONS
         31896 GLYCOSYLATION
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=> mutated and L6
         28228 MUTATED
L13
             1 MUTATED AND L6
=> truncated and 16
         32137 TRUNCATED
L14
            15 TRUNCATED AND L6
=> D L14 IBIB ABS 1-14
L14 ANSWER 1 OF 15 CAPLUS COPYRIGHT 2005 ACS on STN
                         2005:163200 CAPLUS
ACCESSION NUMBER:
                          Purification and application of C-terminally
TITLE:
                          truncated hepatitis C virus El proteins
```

expressed in Escherichia coli

Liu, Jing; Zhu, Li-Xin; Kong, Yu-Ying; Li, Guang-Di; AUTHOR(S):

Wang, Yuan

State Key Laboratory of Molecular Biology, Institute CORPORATE SOURCE: .

of Biochemistry and Cell Biology, Shanghai Institutes for Biological Sciences, Chinese Academy of Sciences,

Shanghai, 200031, Peop. Rep. China

World Journal of Gastroenterology (2005), 11(4), SOURCE:

503-507

CODEN: WJGAF2; ISSN: 1007-9327 World Journal of Gastroenterology

DOCUMENT TYPE: Journal

PUBLISHER:

English LANGUAGE:

AIM: To explore the possibility of expressing hepatitis C virus (

HCV) envelope protein 1 (E1) in Escherichia coli (E

coli) and to test the purified recombinant El proteins for clin. and research applications. METHODS: C-terminally truncated E1 fragments were expressed in E. coli as hexa-histidine-tagged fusion proteins. The expression products were purified under denaturing conditions using immobilized-metal affinity chromatog. Purified E1 proteins were used to immunize rabbits. Rabbit anti-sera thus obtained were reacted with both E. coli- and mammalian cell-expressed E1 glycoproteins as detected by Western blot. RESULTS: Full-length E1 protein proved difficult to express in E. coli. C-terminally truncated E1 was successfully expressed in E. coli as hexa-histidine-tagged recombinant fusion protein and was purified under denaturing conditions on Ni2+-NTA agarose. anti-sera raised against purified recombinant El specifically reacted with mammalian cell-expressed E1 glycoproteins in Western blot. Furthermore, E. coli-derived El protein was able to detect animal antibodies elicited by E1-based DNA immunization. CONCLUSION: These results demonstrate that the prokaryotically expressed El proteins share identical epitopes with eukaryotically expressed El glycoprotein. The E. coli-derived El proteins and corresponding antisera can become useful tools in anti-HCV vaccine research.

THERE ARE 29 CITED REFERENCES AVAILABLE FOR THIS REFERENCE COUNT: 29 RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L14 ANSWER 2 OF 15 CAPLUS COPYRIGHT 2005 ACS on STN

2003:758562 CAPLUS ACCESSION NUMBER:

DOCUMENT NUMBER: 140:25396

CD81-dependent binding of hepatitis C virus E1E2 TITLE:

heterodimers

AUTHOR(S): Cocquerel, Laurence; Kuo, Chiung-Chi; Dubuisson, Jean;

Levy, Shoshana

Department of Medicine/Division of Oncology, Stanford CORPORATE SOURCE:

University Medical Center, Stanford, CA, 94305, USA

Journal of Virology (2003), 77(19), 10677-10683

CODEN: JOVIAM; ISSN: 0022-538X

American Society for Microbiology PUBLISHER:

Journal DOCUMENT TYPE:

SOURCE:

English LANGUAGE: Hepatitis C virus (HCV) is the leading cause of chronic liver

disease worldwide. HCV is also the major cause of mixed cryoglobulinemia, a B-lymphocyte proliferative disorder. experimentation with native viral proteins is not feasible. Truncated versions of recombinant E2 envelope proteins, used as surrogates for viral particles, were shown to bind

specifically to human CD81. However, truncated E2 may not fully mimic the surface of HCV virions because the virus encodes 2 envelope glycoproteins that associate with each other as E1E2 heterodimers. Here we show that E1E2 complexes efficiently bind to CD81 whereas truncated E2 is a weak binder, suggesting that

truncated E2 is probably not the best tool with which to study cellular interactions. To gain better insight into virus-cell interactions, we developed a method by which to isolate E1E2 complexes that are properly folded. We demonstrate that purified E1E2 heterodimers bind to cells in a CD81-dependent manner. Furthermore, engagement of B cells by purified E1E2 heterodimers results in their aggregation and in protein tyrosine phosphorylation, a hallmark of B-cell activation. These studies provide a possible clue to the etiol. of HCV-associated B-cell lymphoproliferative diseases. They also delineate a method by which to isolate biol. functional E1E2 complexes for the study of virus-host cell interaction in other cell types.

60

L14 ANSWER 3 OF 15 CAPLUS COPYRIGHT 2005 ACS on STN ACCESSION NUMBER: 2002:180436 CAPLUS

DOCUMENT NUMBER: 137:227162

REFERENCE COUNT:

CORPORATE SOURCE:

TITLE: Cloning and expression of human CD81 major extracellular loop in E. coli and its activity

AUTHOR(S): Zhang, Guojun; Ling, Shigan; Song, Xiaoguo; Zhang,

Heqiu; Chen, Kun; Zhu, Cuixia; Xiu, Bingshui Institute of Basic Medical Sciences, Academy of

Military Medical Sciences, Beijing, 100850, Peop. Rep.

THERE ARE 60 CITED REFERENCES AVAILABLE FOR THIS

RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

China

SOURCE: Junshi Yixue Kexueyuan Yuankan (2001), 25(4), 260-264

CODEN: JYKYEL; ISSN: 1000-5501

PUBLISHER: Junshi Yixue Kexueyuan Yuankan Bianjibu

DOCUMENT TYPE: Journal LANGUAGE: Chinese

An expression plasmid for a fusion protein of human CD81 major extracellular loop was constructed and binding activity of its expressed protein with HCV E2 was studied. CD81 major extracellular loop sequence was amplified from human peripheral blood lymphocytes by RT-PCR, then inserted into the expression vector pBVIL1, and expressed in E. coli. The purified fusion protein was tested for binding activity with E2. CD81-EC2 gene was correctly amplified and inserted into the vector as confirmed by sequencing. The preliminary study showed that the recombinant CD81/EC2 could bind truncated HCV E2 (384-661) protein expressed in E. coli. This work proved the way for further study on interactions of CD81 with HCV and its E2, and for preparation of anti-EC2 monoclonal antibody.

L14 ANSWER 4 OF 15 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 2001:912910 CAPLUS

DOCUMENT NUMBER: 137:104371

TITLE: Secretory expression of different C-terminal

truncated HCV El proteins in

mammalian cells and characterization of the expressed

products

AUTHOR(S): Zhu, Jun; Kong, Yuying; Liu, Jing; Zhang, Zuchuan;

Wang, Yuan; Li, Guangdi

CORPORATE SOURCE: Institute of Biochemistry and Cell Biology, Shanghai

Institute for Biological Sciences, Chinese Academy of

Sciences, Shanghai, 200031, Peop. Rep. China

SOURCE: Shengwu Huaxue Yu Shengwu Wuli Xuebao (2001), 33(6),

634-640

CODEN: SHWPAU; ISSN: 0582-9879 Shanghai Kexue Jishu Chubanshe

DOCUMENT TYPE: Journal LANGUAGE: Chinese

PUBLISHER:

Three fragments of HCV envelope 1 (E1) with different C-terminal truncation at aa310, aa325, aa340 were cloned into the mammalian expression vector pSecTagB. An epitope in the hepatitis B surface antigen, preS1(21-47), were genetically engineered onto the N-terminus of the recombinant protein and used as an affinity tag for detection and purification. The resulting pSec-preS1-E1t310, pSec-preS1-E1t325, and pSec- preS1-E1t340 were transiently expressed in the HeLa cells and antigenicity, secretory efficiency, and glycosylation type of the recombinant E1 proteins were compared. All of the three recombinant proteins could be detected by both preS1 monoclonal antibody and E1 polyclonal antiserum. The expression products were secreted and highly mannose-type glycosylated, with S1Elt325 being secreted, indicating the influence of the hydrophobic regions on the secretion of the E1 protein. Three CHO cell lines expressing the proteins, S1Elt310, S1Elt325, and S1Elt340, were established and CHO/pSecS1Elt325 was chosen for further study. The secreted S1Elt325

could be enriched from cell culture medium by the preS1 antibody-coupled Sepharose. The glycosylation anal. indicated the lack of complex glycogen even after the El was secreted via Golgi complexes. The established stable cell lines and anti-preS1 affinity method could be utilized to enrich and purify the HCV El expressed in mammalian cells, and may be used for further characterization of this protein.

L14 ANSWER 5 OF 15 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 2000:546735 CAPLUS

133:295002 DOCUMENT NUMBER:

Characterization of Modified Hepatitis C Virus E2 TITLE:

Proteins Expressed on the Cell Surface

Forns, Xavier; Allander, Tobias; Rohwer-Nutter, AUTHOR(S):

Patricia; Bukh, Jens

Hepatitis Viruses Section, National Institutes of CORPORATE SOURCE:

> Health, Bethesda, MD, 20892, USA Virology (2000), 274(1), 75-85 CODEN: VIRLAX; ISSN: 0042-6822

Academic Press PUBLISHER:

Journal DOCUMENT TYPE: English LANGUAGE:

SOURCE:

The envelope proteins of hepatitis C virus (HCV) are AB

the likely targets of neutralizing antibodies and their mol. and functional characterization is relevant for vaccine development. previously showed that surface-expressed E2 is a better immunogen than intracellular E2 and, therefore, we were interested in exploring more efficient ways to present E2 protein on the cell surface. We found that E2 targeted to the cell surface by replacement of its transmembrane domain did not bring E1 to the surface although E1 could be expressed independently on the cell surface if its transmembrane domain was similarly replaced. FACS anal. suggested that E2 expressed on the cell surface acquired its native conformation more efficiently when truncated at aa 661 than when truncated at aa 715. The shorter form of truncated E2 better retained the ability to bind the second extracellular loop (EC2) of CD81, the putative HCV receptor. Interestingly, deletion of the hypervariable region 1 (HVR1) did not perceptibly alter E2 structure; cell-surface forms of E2 lacking the HVR1 remained reactive with conformation-sensitive MAbs and were able to bind recombinant EC2 of CD81. (c) 2000 Academic Press.

37 THERE ARE 37 CITED REFERENCES AVAILABLE FOR THIS REFERENCE COUNT: RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L14 ANSWER 6 OF 15 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER:

2000:494559 CAPLUS 133:221325

DOCUMENT NUMBER: TITLE:

Evaluation of hepatitis C virus glycoprotein E2 for vaccine design: an endoplasmic reticulum-retained

recombinant protein is superior to secreted recombinant protein and DNA-based vaccine

candidates

AUTHOR(S):

SOURCE:

Heile, Jens M.; Fong, Yiu-Lian; Rosa, Domenico; Berger, Kim; Saletti, Giulietta; Campagnoli, Susanna;

Bensi, Giuliano; Capo, Sabrina; Coates, Steve; Crawford, Kevin; Dong, Christine; Wininger, Mark; Baker, Gary; Cousens, Larry; Chien, David; Ng, Philip; Archangel, Phillip; Grandi, Guido; Houghton, Michael; Abrignani, Sergio

CORPORATE SOURCE:

IRIS Research Center, Siena, 53100, Italy Journal of Virology (2000), 74(15), 6885-6892 CODEN: JOVIAM; ISSN: 0022-538X

American Society for Microbiology

PUBLISHER: DOCUMENT TYPE: Journal

LANGUAGE: English

Hepatitis C virus (HCV) is the leading causative agent of blood-borne chronic hepatitis and is the target of intensive vaccine research. The virus genome encodes a number of structural and nonstructural antigens which could be used in a subunit vaccine. The HCV envelope glycoprotein E2 has recently been shown to bind CD81 on human cells and therefore is a prime candidate for inclusion in any such

vaccine. The expts. presented here assessed the optimal form of HCV E2 antigen from the perspective of antibody generation. quality of recombinant E2 protein was evaluated by both the capacity to bind its putative receptor CD81 on human cells and the ability to elicit antibodies that inhibited this binding (NOB antibodies). We show that truncated E2 proteins expressed in mammalian cells bind with high efficiency to human cells and elicit NOB antibodies in guinea pigs only when purified from the core-glycosylated intracellular fraction, whereas the complex-glycosylated secreted fraction does not bind and elicits no NOB antibodies. We also show that carbohydrate moieties are not necessary for E2 binding to human cells and that only the monomeric nonaggregated fraction can bind to CD81. Moreover, comparing recombinant intracellular E2 protein to several E2-encoding DNA vaccines in mice, we found that protein immunization is superior to DNA in both the quantity and quality of the antibody response elicited. Together, our data suggest that to elicit antibodies aimed at blocking HCV binding to CD81 on human cells, the antigen of choice is a mammalian cell-expressed, monomeric E2 protein purified from the intracellular fraction.

REFERENCE COUNT: 52 THERE ARE 52 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L14 ANSWER 7 OF 15 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 1999:290054 CAPLUS

DOCUMENT NUMBER: 131:113495

TITLE: Comparison of secretion of a hepatitis C virus

glycoprotein in Saccharomyces cerevisiae and

Kluyveromyces lactis

AUTHOR(S): Mustilli, Anna Chiara; Izzo, Emanuela; Houghton,

Michael; Galeotti, Cesira L.

CORPORATE SOURCE: Chiron Vaccines, I.R.I.S., Siena, 53100, Italy

SOURCE: Research in Microbiology (1999), 150(3), 179-187

CODEN: RMCREW; ISSN: 0923-2508

PUBLISHER: Editions Scientifiques et Medicales Elsevier

DOCUMENT TYPE: Journal LANGUAGE: English

A C-terminally truncated form of the hepatitis C virus (HCV) putative envelope glycoprotein E2 was expressed in two yeast species, Saccharomyces cerevisiae and Kluyveromyces lactis, using a yeast signal peptide sequence to direct the viral glycoprotein to the endoplasmic reticulum (ER) pathway of secretion. Characterization of secreted E2 showed that the protein is endoglycosidase-H-sensitive in both yeasts. Moreover, in vivo inhibition of glycosylation with tunicamycin prevented secretion of E2 and showed that, of its 11 putative N-linked glycosylation sites, at least eight were core-glycosylated. Anal. of the heterologous glycoprotein by SDS-PAGE under nonreducing conditions and by gel filtration demonstrated the formation of multiple disulfides, which resulted in secretion of heterogeneous aggregates with an average mol. mass of 770-1000 kDa in both yeasts. However, variations were observed in the binding of the glycoprotein secreted by the two yeasts to a mannose-specific lectin, and also in its reactivity with anti-E2-specific antibodies. This denotes differences between the two yeasts in folding and/or modification of the E2 glycoprotein.

REFERENCE COUNT: 32 THERE ARE 32 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L14 ANSWER 8 OF 15 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 1998:810677 CAPLUS

DOCUMENT NUMBER: 130:166917

TITLE: Identification of a domain containing B-cell epitopes

in hepatitis C virus E2 glycoprotein by using mouse

monoclonal antibodies

AUTHOR(S): Woo Lee, Jae; Kim, Kwang-Mi; Jung, Seung-Hye; Lee, Ki

Jeong; Choi, Eung-Chil; Sung, Young-Chul; Kang,

Chang-Yuil

CORPORATE SOURCE: Laboratory of Immunology, College of Pharmacy, Seoul

National University, Seoul, 151-742, S. Korea

SOURCE: Journal of Virology (1999), 73(1), 11-18

CODEN: JOVIAM; ISSN: 0022-538X

PUBLISHER: American Society for Microbiology

DOCUMENT TYPE: LANGUAGE: Journal English

AB Evidence from clin. and exptl. studies of human and chimpanzees suggests

that hepatitis C virus (HCV) envelope glycoprotein E2

is a key antigen for developing a vaccine against HCV infection. To identify B-cell epitopes in HCV E2, six murine monoclonal antibodies (MAbs), CET-1 to -6, specific for HCV E2 protein were

generated by using recombinant proteins containing E2t (a

C-terminally truncated domain of HCV E2 [amino acids

386 to 693] fused to human growth hormone and glycoprotein D). We tested

whether HCV-infected sera were able to inhibit the binding of

CET MAbs to the former fusion protein. Inhibitory activity was observed in most sera tested, which indicated that CET-1 to -6 were similar to anti-E2 antibodies in human sera with respect to the epitope specificity. The

spacial relationship of epitopes on E2 recognized by CET MAbs was determined by

surface plasmon resonance anal. and competitive ELISA. The data indicated that three overlapping epitopes were recognized by CET-1 to -6. For

mapping the epitopes recognized by CET MAbs, we analyzed the reactivities of CET MAbs to six **truncated** forms and two chimeric forms of

recombinant E2 proteins. The data suggest that the epitopes
recognized by CET-1 to -6 are located in a small domain of E2 spanning

amino acid residues 528 to 546.

REFERENCE COUNT: 46 THERE ARE 46 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L14 ANSWER 9 OF 15 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 1998:592014 CAPLUS

DOCUMENT NUMBER: 129:301407

TITLE: Hepatitis C virus envelope DNA-based

immunization elicits humoral and cellular immune

responses

AUTHOR(S): Lee, Seung Woo; Cho, Jae Ho; Lee, Ki Jeong; Sung,

Young Chul

CORPORATE SOURCE: Department of Life Science, Center for Biofunctional

Molecules, School of Environmental Engineering, Pohang University of Science and Technology, Pohang, 790-784,

S. Korea

SOURCE: Molecules and Cells (1998), 8(4), 444-451

CODEN: MOCEEK; ISSN: 1016-8478

PUBLISHER: Springer-Verlag Singapore Pte. Ltd.

DOCUMENT TYPE: Journal LANGUAGE: English

AB The vaccine development for hepatitis C virus (HCV) is highly

urgent to prevent non A and non B hepatitis. It was recently shown that

the HCV envelope proteins appeared to the key viral

antigens to induce protective immunity. To generate immune responses to

the HCV envelope proteins on the DNA-based

immunization, various **envelope** gene-containing plasmids were constructed. For efficient expression and secretion of **envelope** proteins, the signal sequence of each **envelope** protein was

replaced with either herpes simplex virus type-1 (HSV-1) gD or signal

sequence of gD and truncated C-terminal hydrophobic regions of

envelope proteins. The i.m. injection of these plasmids generated
a significant level of antibody titers to the E1 and E2 proteins, which

maximally reached 850 and 25,000 resp. The secreted form of each **envelope** protein and the fusion of the highly immunogenic gD

proteins were shown to have no significant effect on generating immune

responses to the envelope proteins. In addition, immunized rats

appeared to generate antibodies directed to the homologous HVR-1 peptide. Splenic lymphocytes from immunized rats were shown to induce significant

T-cell proliferative responses with the stimulation of recombinant

E1 and E2 proteins. Our results demonstrated that the **HCV** envelope-DNA based immunization could elicit both humoral and

cellular immune responses.

REFERENCE COUNT: 46 THERE ARE 46 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

1998:313394 CAPLUS ACCESSION NUMBER:

DOCUMENT NUMBER: 129:107767

Isolation and characterization of human monoclonal TITLE:

antibodies against hepatitis C virus envelope

glycoproteins

AUTHOR(S): Da Silva Cardoso, Marcia; Siemoneit, Karl; Sturm,

Daniela; Krone, Christoph; Moradpour, Darius; Kubanek,

Bernhard

CORPORATE SOURCE: Blood Transfusion Service of Baden-Wurttemberg and

Department of Transfusion Medicine, University of Ulm,

Germany

Journal of Medical Virology (1998), 55(1), 28-34 SOURCE:

CODEN: JMVIDB; ISSN: 0146-6615

Wiley-Liss, Inc. PUBLISHER:

DOCUMENT TYPE: Journal English LANGUAGE:

The isolation and characterization of human monoclonal antibodies (humAbs)

against the hepatitis C virus (HCV) glycoproteins E1 and E2 are

described. B-cells from blood donors with anti-HCV were

transformed with Epstein-Barr virus. The supernatants of the resulting lymphoblastoid clones were screened by ELISA with an extract of cells infected with a recombinant vaccinia virus RMPA95 expressing the

envelope proteins E1 and E2 of an HCV genotype 1a virus

(H strain). Pos. clones were fused to the heteromyeloma cell line K6H6/B5. Fifteen heterohybridoma cell lines have been established. specificity of the isolated humAbs was determined both by ELISA and Western blot assays. Several recombinant exts. expressing either the E1 or E2 protein or truncated forms were used in an attempt to map

the epitopes on the viral glycoproteins. Some of the humAbs were used successfully for immunofluorescence investigation of transfected cells.

Seven specific anti-E2 humAbs, which react with the envelope protein 2 of genotype 1a and 1b isolates, were characterized.

THERE ARE 22 CITED REFERENCES AVAILABLE FOR THIS REFERENCE COUNT: 22

RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L14 ANSWER 11 OF 15 CAPLUS COPYRIGHT 2005 ACS on STN

1997:775256 CAPLUS ACCESSION NUMBER:

DOCUMENT NUMBER: 128:72872

Hepatitis C virus E2 protein purified from mammalian TITLE:

cells is frequently recognized by E2-specific

antibodies in patient sera

Lee, Ki Jeong; Suh, Young-Ah; Cho, Young Gyu; Cho, AUTHOR(S):

> Young Shik; Ha, Gun Woo; Chung, Kwang-Hoe; Hwang, Jae Hoon; Yun, Young Dae; Lee, Dong Soon; Kim, Chang Min; Sung, Young-Chul

CORPORATE SOURCE: Department of Life Science, Center for Biofunctional

Molecules, School of Environmental Engineering, Pohang University of Science and Technology, Pohang, 790-784,

S. Korea

SOURCE: Journal of Biological Chemistry (1997), 272(48),

30040-30046

CODEN: JBCHA3; ISSN: 0021-9258

PUBLISHER: American Society for Biochemistry and Molecular

Biology

DOCUMENT TYPE: Journal LANGUAGE: English

AΒ The envelope protein of hepatitis C virus (HCV) is

composed of two membrane-associated glycoproteins, E1 and E2. HCV E2 protein as a secretory form at a high level, we constructed

a recombinant chinese hamster ovary (CHO) cell line expressing a C-terminal truncated E2 (E2t) fused to human growth hormone

(hGH), CHO/hGHE2t. The hGHE2t fusion protein was purified from the culture supernatant using anti-hGH mAb affinity chromatog. at approx. 80% purity. The purified hGHE2t protein appeared to be assembled into oligomers linked by intermol. disulfide bond(s) when d. gradient centrifugation and SDS-polyacrylamide gel electrophoresis were employed.

When the purified fusion protein was used for testing its ability to bind

to antibodies specific for HCV by ELISA, the protein was

recognized by antibodies in sera from 90% of HCV-pos. patients.

Treatment of hGHE2t protein by β -mercaptoethanol, but not by heat and SDS, significantly reduced its reactivity to the antibodies of patient sera, suggesting that intermol. and/or intramol. disulfide bonds are important for its ability to recognize its specific antibody and that the E2 protein contains discontinuous antigenic epitope(s).

REFERENCE COUNT: 38 THERE ARE 38 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L14 ANSWER 12 OF 15 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 1997:113448 CAPLUS

DOCUMENT NUMBER: 126:117059

TITLE: Method for detection of antibody to hepatitis C virus

second envelope glycoprotein

INVENTOR(S): Okasinski, Gregory F.; Schaefer, Verlyn G.; Suhar,

Thomas S.; Lesniewski, Richard R.; Scheffel, James W.

PATENT ASSIGNEE(S): Abbott Laboratories, USA SOURCE: PCT Int. Appl., 34 pp.

CODEN: PIXXD2

DOCUMENT TYPE: Patent LANGUAGE: English

FAMILY ACC. NUM. COUNT: 1

PATENT INFORMATION:

PATENT NO.	KIND DATE		APPLICATION NO.	DATE				
WO 9641196	A1	19961219	WO 1996-US8536	19960604				
· · · · · · · · · · · · · · · · · · ·			FR, GB, GR, IE, IT,					
CA 2223277 EP 836708	AA A1		CA 1996-2223277 EP 1996-917969	19960604 19960604				
R: AT, BE, CH,	DE, ES	, FR, GB, I	IT, LI, NL					
01 1100.110	Т2	19990622	JP 1996-501105	19960604				
PRIORITY APPLN. INFO.:			US 1995-481018 WO 1996-US8536	A 19950607 W 19960604				

AB A method for detecting antibody to HCV in a test sample. The method includes utilizing a recombinant protein that is the expression product of mammalian cells transformed by a heterologous expression vector comprising a DNA sequencing encoding an E2 truncated protein. Test kits which include this recombinant protein also are provided.

L14 ANSWER 13 OF 15 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 1996:698698 CAPLUS

DOCUMENT NUMBER: 126:6277

TITLE: Expression of HCV envelope

proteins and the serological utility of the anti-E2

immune response

AUTHOR(S): Lesniewski, Richard R.; Watanabe, Shinichi; Devare,

Sushil G.

CORPORATE SOURCE: Hepatitis Research and Development, Abbott

Laboratories, Abbott Park, IL, 60064, USA

SOURCE: Proceedings of the International Symposium of the

Princess Takamatsu Cancer Research Fund (1995), Volume Date 1994, 25th (Hepatitis C Virus and Its Involvement

in the Development of Hepatocellular Carcinoma),

129-137

CODEN: PPTCBY

PUBLISHER: Princeton Scientific

DOCUMENT TYPE: Journal LANGUAGE: English

AB The 5' end of the hepatitis C virus (HCV) genome encodes structural proteins of the virion. The first gene encodes a highly basic core protein. Immediately downstream of the core gene are regions which encode the envelope proteins (E1 and E2) of the virus.

Artificial expression and secretion of immunol. active envelope proteins have proven to be a substantial challenge due to the high degree of glycosylation and the existence of certain hydrophobic domains contained within these sequences. Bacterial cell expression of recombinant HCV envelope proteins results in

products that are not glycosylated and are poorly immunogenic. Emphasis has shifted to the use of mammalian cell lines (human embryonic kidney [HEK] and Chinese hamster ovary [CHO] cells) for the expression of glycosylated, immunol. active envelope proteins. Using HEK cells, E1 is expressed intracellularly but is not secreted from the cells. When El is cloned in fusion with a C-terminal truncated E2 protein, both proteins are detected intracellularly; however, only E2 is secreted. When the E1/E2 processing site is interrupted by constructing deletion mutants, the unprocessed E1/E2 fusion protein can be secreted from the cells. Quantifiable expression and secretion of a truncated E2 protein is now possible using CHO cells and SV40-based vectors. The HCV E2 glycoprotein expressed from CHO cells is highly antigenic; a strong humoral response to this antigen develops in persons infected with HCV. Antibodies to E2 are found in 95% of patients with detectable HCV RNA in their sera. The presence of antibodies to E2 is not indicative of viral clearance and therefore the role these antibodies play in protective immunity, if any, is unclear.

L14 ANSWER 14 OF 15 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 1996:260624 CAPLUS

DOCUMENT NUMBER: 124:312065

TITLE: Processing of the El glycoprotein of hepatitis C virus

expressed in mammalian cells

AUTHOR(S): Fournillier-Jacob, Anne; Cahour, Annie; Escriou,

Nicolas; Girard, Marc; Wychowski, Czeslaw

CORPORATE SOURCE: Institut Pasteur, Unite Virologie Moleculaire, Paris,

75724, Fr.

SOURCE: Journal of General Virology (1996), 77(5), 1055-64

CODEN: JGVIAY; ISSN: 0022-1317 Society for General Microbiology

DOCUMENT TYPE: Journal LANGUAGE: English

PUBLISHER:

The structural part of the hepatitis C virus (HCV) genome AΒ encodes a capsid protein, C, and two envelope glycoproteins, E1 and E2, released from the virus polyprotein precursor by signalase(s) cleavage(s). The processing of El was investigated by infecting simian cells with recombinant vaccinia viruses expressing parts of the HCV structural proteins. When the predicted El sequence was expressed alone (amino acid residues 174-370 of the polyprotein) or with the capsid protein gene (residues 1-370), it showed an apparent mol. mass of 35 kDa as measured by SDS-PAGE anal. However, when El was expressed as part of a truncated C-E1-truncated E2 polypeptide (residues 132-383), the processed El product had the expected apparent mol. mass of 31 kDa, suggesting that flanking sequences are necessary for the generation of the mature 31 kDa El form. The N-terminal sequence of the two El forms was found to be the same. Anal. of the glycosylation pattern showed that, in both species, only four of the five potential N-linked glycosylation sites were recognized, indicating that glycosylation was not involved in the mol. mass difference. We showed that expression of El with or without the hydrophobic stretch of amino acids residues 371-383, defined as the E2 signal sequence, may be responsible for the difference in electrophoretic mobility of the two E1 species. In vitro translation assays and site-directed mutagenesis expts. suggest that this sequence remains part of the 31 kDa El mature protein.

=> D L14 IBIB abs 15

L14 ANSWER 15 OF 15 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 1994:698202 CAPLUS

DOCUMENT NUMBER: 121:298202

TITLE: Processing of E1 and E2 glycoproteins of hepatitis C

virus expressed in mammalian and insect cells

AUTHOR(S): Matsuura, Yoshiharu; Suzuki, Tetsuro; Suzuki, Ryosuke;

Sato, Mitsuru; Aizaki, Hideki; Saito, Izumu; Miyamura,

Tatsuo

CORPORATE SOURCE: Dep. Virology II, Natl. Inst. Health, Tokyo, 162,

Japan

SOURCE: Virology (1994), 205(1), 141-50 CODEN: VIRLAX; ISSN: 0042-6822

PUBLISHER: Academic
DOCUMENT TYPE: Journal
LANGUAGE: English

Processing of the envelope glycoproteins (E1 and E2) of hepatitis C virus (HCV) was investigated by using cDNA clones covering the structural and part of the nonstructural (NS) protein regions. The cDNA clones expressed in mammalian and insect cells were immunopptd. by serum of a hepatitis C patient and by monoclonal and polyclonal antibodies riased against the recombinant proteins expressed in insect cells or Escherichia coli. The E2 protein expressed in both insect and mammalian cells was a glycoprotein of 60 kDa (gp60) and removal of the sugar residues by N-glycanase yielded 38- and 40-kDa proteins. Pulse-chase expts. revealed that efficient expression and processing of the envelope proteins required coexpression with the flanking core and NS2 proteins. Not only E1 and E2 proteins but also NS2 and NS3 proteins were copptd. by anti-E1 or anti-E2 monoclonal antibody in the cells infected with the recombinant baculovirus expressing structural and NS proteins (NS2 and NS3), while only the NS3 protein was precipitated by anti-NS3 antibody. The association of E1 and E2 proteins was not influenced by the presence of a reducing agent and was still observed in the cells coinfected with the deletion mutants lacking both internal and C-terminal hydrophobic regions of each protein. Furthermore, the truncated forms of the E1 and E2 proteins were secreted into the culture supernatant and some of them were still associated with each other.

=> D L13 IBIB ABS

L13 ANSWER 1 OF 1 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER:

2001:338732 CAPLUS

DOCUMENT NUMBER:

134:352270

TITLE:

Fusion proteins containing antigenic ectodomain of measles virus hemagglutin protein and viral targeting

peptides and its use as vaccines

INVENTOR(S):

Petrik, Juraj

PATENT ASSIGNEE(S):

UK

SOURCE:

PCT Int. Appl., 27 pp.

CODEN: PIXXD2

DOCUMENT TYPE:

TYPE: Patent

LANGUAGE: English

FAMILY ACC. NUM. COUNT:

PATENT INFORMATION:

	PATENT NO.					KIND DATE			APPLICATION NO.					DATE					
	WO					A1	_	2001	0510							20001101			
		W:	ΑE,	AG,	AL,	AM,	AT,	AU,	AZ,	BA,	BB,	BG,	BR,	BY,	ΒZ,	CA,	CH,	CN,	
			CR,	CU,	CZ,	DE,	DK,	DM,	DZ,	EE,	ES,	FI,	GB,	GD,	GE,	GH,	GM,	HR,	
			HU,	ID,	IL,	IN,	IS,	JP,	ΚE,	KG,	ΚP,	KR,	ΚZ,	LC,	LK,	LR,	LS,	LT,	
			LU,	LV,	MA,	MD,	MG,	MK,	MN,	MW,	MX,	MZ,	NO,	NZ,	PL,	PT,	RO,	RU,	
			SD,	SE,	SG,	SI,	SK,	SL,	TJ,	TM,	TR,	TT,	ΤZ,	UA,	UG,	US,	UZ,	VN,	
			YU,	ZA,	ZW,	AM,	ΑZ,	BY,	KG,	KZ,	MD,	RU,	ТJ,	MT					
		RW:	GH,	GM,	ΚĖ,	LS,	MW,	MZ,	SD,	SL,	SZ,	TZ,	UG,	ZW,	AT,	BE,	CH,	CY,	
			DE,	DK,	ES,	FI,	FR,	GB,	GR,	ΙE,	IT,	LU,	MC,	NL,	PT,	SE,	TR,	BF,	
			ВJ,	CF,	CG,	CI,	CM,	GΑ,	GN,	GW,	ML,	MR,	NE,	SN,	TD,	TG			
		2389																	
		2001																	
	ΕP	1226																	
		R:	ΑT,	BE,	CH,	DE,	DK,	ES,	FR,	GB,	GR,	IT,	LI,	LU,	NL,	SE,	MC,	PT,	
								RO,											
		2003																	
PRIO	PRIORITY APPLN. INFO.:				. :						GB 1					_	9991		
									WO 2000-GB4191					W 20001101					
AB	AB A recombinant bifu						nctional fusion or			rotein comprises a fi					irst				

AB A recombinant bifunctional fusion protein comprises a first component which is a mutated antigenic ectodomain of measles virus hemagglutin protein (MeaH); and a second component fused thereto which is capable of binding to the surface structure of genetically

variable viruses such as HCV or HIV or other therapeutic targets. The MeaH antigenic ectodomain is genetically modified and does not to bind to CD46 receptor or cause hemadsorption or hemagglutination, but retains its antigenicity and is recognized by anti-measles antibodies, thus it serves as booster/carrier antigen. The second component binds to the target and the first component is recognized by anti-measles antibodies present in the majority of the population. Examples of second component for HIV gp120env targeting are provided by screening human expression cDNA library with biotinylated recombinant env. protein may be used therapeutically to treat HCV or HIV infection or against other therapeutic targets.

REFERENCE COUNT:

THERE ARE 2 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

=> D L12 IBIB ABS 1-5

L12 ANSWER 1 OF 5 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 2002:832824 CAPLUS

DOCUMENT NUMBER:

137:351491

TITLE:

Production of recombinant HCV envelope proteins with expression

vectors encoding avian lysozyme leader or

signal peptide

INVENTOR(S):

Sablon, Erwin; Van Broekhoven, Annie; Bosman, Alfons;

Depla, Erik; Deschamps, Geert

PATENT ASSIGNEE(S):

Innogenetics N.V., Belg.

SOURCE:

PCT Int. Appl., 319 pp.

CODEN: PIXXD2

DOCUMENT TYPE:

Patent

LANGUAGE:

English

FAMILY ACC. NUM. COUNT:

PATENT INFORMATION:

PA'	PATENT NO.				KIND DATE			APPLICATION NO.						DATE			
	2002 2002				A2 A3					WO 2	2002-1	BE62			20020424		
	w:	AE, CO, GM, LS, PL, UA,	AG, CR, HR, LT, PT, UG,	AL, CU, HU, LU, RO,	AM, CZ, ID, LV, RU,	AT, DE, IL, MA, SD,	AU, DK, IN, MD, SE,	AZ, DM, IS, MG, SG,	BA, DZ, JP, MK, SI,	EC, KE, MN, SK,	BG, EE, KG, MW, SL,	ES, KP, MX, TJ,	FI, KR, MZ, TM,	GB, KZ, NO, TN,	GD, LC, NZ, TR,	GE, LK, OM, TT,	GH, LR, PH, TZ,
CA	RW:	CY, BF,	GM, DE, BJ,	DK, CF,	ES,	FI, CI,	FR, CM,	GB, GA,	GR, GN,	ΙΕ, GQ,	TZ, IT, GW, 2002-	LU, ML,	MC, MR,	NL, NE,	PT, SN,	SE,	TR, TG
US US US	2003 2003 2003 1381	1085 1529 2115	61 40 97		A1 A1		2003 2003 2003	0612 0814 1113		US 2 US 2 US 2	2002- 2002- 2002- 2002-	1285 1285 1285	90 87 78		2 2 2	0020 0020	424 424 424
		AT, IE,	BE, SI,	CH, LT,	DE, LV,	DK, FI,		FR, MK,	GB, CY,	GR, AL,	IT,	LI,	LU,		SE,		PT,
JP BR ZA	2004 2002 2003	5365 0090 0082	82 33 77		T2 A A		2004 2005 2004	1209 0111 0708		JP 2 BR 2 ZA 2	2002- 2002- 2003-	5834 9033 8277	58		2 2 2	0020 0020 0031	424 424 023
ZA BG	ZA 2003008272 ZA 2003008274 BG 108373 CORITY APPLN. INFO.:						2005 2005 2004	0124		ZA 2 BG 2	2003-1 2003-1 2003-1 2001-1	8274 1083	73		2 2	0031 0031 0031 0010	023 121
	50)								1	WO 2	2001-1 2002-1	BE62		Ţ	_	0010 0020	

AB The current invention relates to vectors and methods for efficient expression of HCV envelope proteins in eukaryotic cells. More particularly said vectors comprise the coding sequence for an avian lysozyme signal peptide or a functional equivalent thereof joined to a **HCV** envelope protein or a part thereof. Said avian lysozyme signal peptide is efficiently removed when the protein comprising said avian lysozyme signal peptide joined to a **HCV** envelope protein or a part thereof is expressed in a eukaryotic cell. Suitable eukaryotic cells include yeast cells such as Saccharomyces or Hansenula cells.

L12 ANSWER 2 OF 5 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 2001:912910 CAPLUS

DOCUMENT NUMBER: 137:104371

TITLE: Secretory expression of different C-terminal truncated

HCV El proteins in mammalian cells and

characterization of the expressed products

AUTHOR(S): Zhu, Jun; Kong, Yuying; Liu, Jing; Zhang, Zuchuan;

Wang, Yuan; Li, Guangdi

CORPORATE SOURCE: Institute of Biochemistry and Cell Biology, Shanghai

Institute for Biological Sciences, Chinese Academy of

Sciences, Shanghai, 200031, Peop. Rep. China

Shengwu Huaxue Yu Shengwu Wuli Xuebao (2001), 33(6),

634-640

CODEN: SHWPAU; ISSN: 0582-9879 Shanghai Kexue Jishu Chubanshe

DOCUMENT TYPE: Journal LANGUAGE: Chinese

SOURCE:

PUBLISHER:

AB Three fragments of HCV envelope 1 (E1) with different

C-terminal truncation at aa310, aa325, aa340 were cloned into the

mammalian expression vector pSecTagB. An epitope in the

hepatitis B surface antigen, preS1(21-47), were genetically engineered

onto the N-terminus of the **recombinant** protein and used as an

affinity tag for detection and purification The resulting pSec-preS1-E1t310,

pSec-preS1-Elt325, and pSec- preS1-Elt340 were transiently expressed in

the HeLa cells and antigenicity, secretory efficiency, and glycosylation type of the recombinant El proteins were compared. All of the three recombinant proteins could be detected by both preSl monoclonal antibody and El polyclonal antiserum.

The expression products were secreted and highly mannose-type glycosylated, with S1E1t325 being secreted, indicating the influence of the hydrophobic regions on the secretion of the E1 protein. Three CHO cell lines expressing the proteins, S1E1t310, S1E1t325, and S1E1t340, were

established and CHO/pSecS1Elt325 was chosen for further study. The secreted S1Elt325 could be enriched from cell culture medium by the preS1 antibody-coupled Sepharose. The glycosylation anal. indicated

the lack of complex glycogen even after the El was secreted via Golgi complexes. The established stable cell lines and anti-preSl affinity

method could be utilized to enrich and purify the HCV E1

expressed in mammalian cells, and may be used for further characterization of this protein.

L12 ANSWER 3 OF 5 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 1996:698698 CAPLUS

DOCUMENT NUMBER: 126:6277

TITLE: Expression of HCV envelope

proteins and the serological utility of the anti-E2

immune response

AUTHOR(S): Lesniewski, Richard R.; Watanabe, Shinichi; Devare,

Sushil G.

CORPORATE SOURCE: Hepatitis Research and Development, Abbott

Laboratories, Abbott Park, IL, 60064, USA

SOURCE: Proceedings of the International Symposium of the

Princess Takamatsu Cancer Research Fund (1995), Volume Date 1994, 25th (Hepatitis C Virus and Its Involvement

in the Development of Hepatocellular Carcinoma),

129-137

CODEN: PPTCBY

PUBLISHER: Princeton Scientific

DOCUMENT TYPE: Journal LANGUAGE: English

AB The 5' end of the hepatitis C virus (HCV) genome encodes

structural proteins of the virion. The first gene encodes a highly basic

core protein. Immediately downstream of the core gene are regions which encode the envelope proteins (E1 and E2) of the virus. Artificial expression and secretion of immunol. active envelope proteins have proven to be a substantial challenge due to the high degree of glycosylation and the existence of certain hydrophobic domains contained within these sequences. Bacterial cell expression of recombinant HCV envelope proteins results in products that are not glycosylated and are poorly immunogenic. Emphasis has shifted to the use of mammalian cell lines (human embryonic kidney [HEK] and Chinese hamster ovary [CHO] cells) for the expression of qlycosylated, immunol. active envelope proteins. Using HEK cells, El is expressed intracellularly but is not secreted from the cells. When E1 is cloned in fusion with a C-terminal truncated E2 protein, both proteins are detected intracellularly; however, only E2 is secreted. the E1/E2 processing site is interrupted by constructing deletion mutants, the unprocessed E1/E2 fusion protein can be secreted from the cells. Quantifiable expression and secretion of a truncated E2 protein is now possible using CHO cells and SV40-based vectors. HCV E2 glycoprotein expressed from CHO cells is highly antigenic; a strong humoral response to this antigen develops in persons infected with HCV. Antibodies to E2 are found in 95% of patients with detectable HCV RNA in their sera. The presence of antibodies to E2 is not indicative of viral clearance and therefore the role these antibodies play in protective immunity, if any, is unclear.

L12 ANSWER 4 OF 5 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 1996:295079 CAPLUS

DOCUMENT NUMBER: 124:352673

TITLE: Recombinant production and purification of

hepatitis C virus envelope proteins for

diagnostic and therapeutic use

INVENTOR(S): Maertens, Geert; Bosman, Fons; De Martynoff, Guy;

Buyse, Marie-Ange

PATENT ASSIGNEE(S): Innogenetics N.V., Belg.

SOURCE: PCT Int. Appl., 146 pp.

CODEN: PIXXD2

DOCUMENT TYPE: Patent LANGUAGE: English

EANGUAGE: English FAMILY ACC. NUM. COUNT: 6

PATENT INFORMATION:

PA	rent n	10.			KIN	KIND DATE			APPLICATION NO.						DATE			
	96043 96043									WO 1	995-1	EP30	31		1	9950	731	
	W:						BR,											
			MN,				KE, NZ,											
	RW:	ΚE,	MW,				AT,											
			MC, TD,		PT,	SE,	BF,	ВJ,	CF,	CG,	CI,	CM,	GA,	GN,	ML,	MR,	NE,	
CA	21722	273	10,	10	AA		1996	0215		CA 1	995-	2172	273		1:	9950	731	
ΑU	95338	324			A1		19960	0304		AU 1	995-	3382	4		1:	9950	731	
	70817																	
ΕP	72150 72150)5			A1		1996	0717		EP 1	995-	9304	34		1	9950	731	
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JP	09503 95060	3396			Т2		1997	0408	1	JP 1	995-	5061	89		1:	9950	731	
BR	95060)59			Α		1997	1028		BR 1	995-	6059			1:	9950	731	
SG	71728 21734	3			A1		2000	0418		SG 1	997-	3877			1:	9950	/31	
					E		2002	0515		AT 1	995-	9304	34		1	9950	731	
	12113						2002											
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EC.	72150)5)57			m o		2002.	1116		PT 1	995-	0204	34 34		1	9950	/31 721	
110	61501	37			12		2002. 2000:	1121		11G 1	995-	5304. 6120'	34 73		1	9950	/ J L	
	62455				A R1		2001	1121		110 1	990-	9275	13 97		1	330U. 3370	011	
	68907				R1		2005	0510		IIS 1	997-	9287	57		1	9 <i>91</i> 0	912	
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	AU 757962			B2			0313	AU	1999-57127		19991029
	AU 995712			A1			0217		2001 000203		20010706
	US 200303			A1			0220		2001-899303		
	US 200218			A1	_		1205		2001-973025		20011010
	US 200309	95980		A1	2	2003	0522	US	2001-995808		20011129
	JP 200422	2729		A2	2	2004	0812	JF	2004-51709		20040226
	US 200418	35061		A1	2	2004	0923	US	2004-825219		20040416
PRIOF	RITY APPLN	I. INFO.:						EF	1994-870132	Α	19940729
•								EF	1994-EP94870132	Α	19940729
								EF	1995-930434	A3	19950731
								JF	1996-506189	A3	19950731
								WC	1995-EP3031	W	19950731
								-	1996-612973	A3	19960311
									1997-928017	В3	19970911
								EF		A	19980624
								EF		A	19990222
								WC		W	19990623
								US		W	19990723
								EF		A	19991027
											19991207
								US		A1	
								US		P	20001201
								US		P	20010111
								US		P	20010830
									2001-973025	A2	20011010
AB	Envelope	proteins	E1	and	E2	of	hepati	itis	C virus (HCV		

Envelope proteins El and E2), their recombinant production and purification, their fragments and engineered derivs., their antigenic epitope peptides, their monoclonal antibodies, and their use for diagnostic and therapeutic means are provided. A method is described for purifying recombinant HCV single or specific oligomeric envelope proteins, characterized in that upon lysing the transformed host cells to isolate the recombinantly expressed protein a disulfide bond cleavage or reduction step is carried out with a disulfide bond cleavage agent (such as dithiothreitol and/or Empigen BB) and an SH group protecting agent (such as N-ethylmaleimide). Various forms of the El and E2 proteins are constructed by standard genetic techniques using vaccinia virus recombination vectors; such proteins are specific for various HCV genotypes, may delete the hydrophobic region from E1, or remove various glycosylation sites; they may also add factor Xa cleavage sites and His6 tags for improved purification Epitope (such as F, G, H, and I) peptides are used to generate monoclonal antibodies and to monitor disease progression in patients. Furthermore, the HCV El protein and peptides are used for prognosing and monitoring the clin. effectiveness and/or clin. outcome of HCV treatment.

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L12 ANSWER 5 OF 5 CAPLUS COPYRIGHT 2005 ACS on STN
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ACCESSION NUMBER:

1992:528131 CAPLUS

DOCUMENT NUMBER:

117:128131

TITLE:

Hepatitis C virus asialoglycoproteins manufacture for

vaccines or immunoassay

INVENTOR(S):

Ralston, Robert O.; Marcus, Frank; Thudium, Kent B.;

Gervase, Barbara A.; Hall, John A.

PATENT ASSIGNEE(S):

Chiron Corp., USA

SOURCE:

PCT Int. Appl., 28 pp.

CODEN: PIXXD2

DOCUMENT TYPE:

Patent

LANGUAGE:

English

FAMILY ACC. NUM. COUNT:

PATENT INFORMATION:

PATENT NO.	KIND DATE	APPLICATION NO.	DATE
WO 9208734	71 10020520	WO 1991-US8272	19911107
			19911107
	FI, HU, JP, NO,		
RW: AT, BE, CH,	DE, DK, ES, FR,	GB, GR, IT, LU, NL, SE	
EP 414475	A1 19910227	EP 1990-309120	19900821
EP 414475	B1 19971210		
R: AT, BE, CH,	DE, DK, ES, FR,	GB, GR, IT, LI, LU, NL,	SE
AT 161041		AT 1990-309120	

ES	2110411 2064705 2064705 9102820		Т3	19980216	ES 1990-309120		19900821	
	2064705		AA	19910226			19900822	
	2064705		C	19990406				
	0102020		7.1	10010307			19900822	
WO	9102820		AI	19910307	WO 1990-054766		19900022	
	W: AU, CA,	JP						
ΑU	9063449		A1	19910403	AU 1990-63449 JP 1990-512531 JP 2001-75114		19900822	
ΑU	655156		В2	19941208				
JP	05502156		Т2	19930422	JP 1990-512531		19900822	,
.TD	2001314192		Δ2	20011113	TP 2001-75114		19900822	
MO	9115771		7.1	10011017	WO 1991-US2225		10010320	
WO								
				CA, FI, GB,	HU, JP, KP, KR, LK,	MC,	MG, MW, NO,	
	PL, RO,							
	RW: BF, BJ,	CF,	CG,	CM, GA, ML,	MR, SN, TD, TG			
ΑU	9176510		A1	19911030	AU 1991-76510		19910329	
Δ[]	639560		B2	19930729				
CD	2257701		7.1	10030120	.CB 1002-20490		10010320	
25	0100000		Ϋ́	19930120	DD 1001 6200		10010329	
BR	9106309		A	19930420	BR 1991-6309		19910329	
ΗU	62706		A2	19930528	HU 1992-3146		19910329	
ΗU	217025		В	19991129				
JΡ	05508219		Т2	19931118	JP 1991-507636		19910329	
.TP	2733138		R2	19980330				•
BO.	100016		D 1	19950330	PO 1975-92012		10010320	
NO.	170100		D1	10070020	NO 1973-92012		10010329	
PL	1/2133		BI	19970829	PL 1991-296329		19910329	
RU	2130969		C1	19990527	RU 1991-5053084		19910329	
EΡ	450931		A1	19911009	MR, SN, TD, TG AU 1991-76510 GB 1992-20480 BR 1991-6309 HU 1992-3146 JP 1991-507636 RO 1975-92012 PL 1991-296329 RU 1991-5053084 EP 1991-302910		19910403	
EΡ	450931		В1	19960612				
					GB, GR, IT, LI, LU,	NT.	SE	
FD	693687	,	Δ1	19960124	FP 1995-114016	,	19910403	
ED.	603607		D1	10000720	EP 1995-114016		17710403	
EP								
	R: AT, BE,	CH,	DE,	DK, ES, FR,	GB, GR, IT, LI, LU,	NΓ,	SE	
ΑT	139343		E	19960615	AT 1991-302910		19910403	
ES	2088465		Т3	19960816	ES 1991-302910		19910403	
ΑТ	182684		E	19990815	AT 1991-302910 ES 1991-302910 AT 1995-114016 ES 1995-114016 CA 1991-2095521 AU 1991-90267 EP 1992-900091		19910403	
ES	2134388		ጥጓ	19991001	ES 1995-114016		19910403	
C Z	2005521		ממ	10020500	CN 1001 2005521		10011107	
CA	2093321		AA	19920509	CA 1991-2095521		19911107	
ΑU	9190267		ΑI	19920611	AU 1991-90267		19911107	
ΑU	668078		В2	19960426				
EΡ	556292		A1	19930825	EP 1992-900091		19911107	
EΡ	556292		В1	19991229				
					GB, GR, IT, LI, LU,	NT.	SE	
.TD	06504431	0117	T2	19940526	TD 1992-500944			
	00004401		7.2	19940526	JP 1992-500944		10011107	
	66063		AZ	19940926	UO 1993-1336			
	842947		A2				19911107	
EΡ	842947		А3					
EΡ	842947		В1	20040421				
	R: AT, BE,	CH.	DE.		GB, GR, IT, LI, LU,	NL.	SE	
RU	2123528	•	C1	19981220		•	19911107	
	175610		B1	19990129			19911107	•
	188220		E	20000115			19911107	
	2139591		Т3	20000216			19911107	
	115446		В1	20000228	RO 1993-626		19911107	
CA	2203443		С	20010828	CA 1991-2203443		19911107	
JP	2001286290		A2	20011016	JP 2001-59335		19911107	
	289006		В6	20011017			19911107	•
	2175657		C2	20011110			19911107	
	2003093081			20030402			19911107	•
			A2					
	2003174875		A2	20030624			19911107	
	1471073		A2	20041027			19911107	
ΕP	1471073		A3	20041201				
	R: AT, BE,	CH,	DE,	DK, ES, FR,	GB, GR, IT, LI, LU,	NL,	SE	
FI	106317	•	В1	20010115		•	19920928	
	9203839		A	19921119			19921001	
	310241		B1	20010611			17721001	
							10020505	
	107803		В1	20011015			19930505	
	9301680		Α	19930628			19930507	
	304380		В1	19981207				
$rac{\Gamma}{\Lambda}$	10344		В	19960220	LV 1993-4381		19930531	
	5679342		Α	19971021			19930727	
	3808		В	19960325			19931230	
			_					

US 5968775 US 5712087 US 6312889 FI 9701702	A A B1 A	19991019 19980127 20011106 19970421	US US	1995-438435 1995-440519 1995-440549 1997-1702		19950510 19950512 19950512 19970421
FI 107804 NO 9702213	B1 A B1	20011015 19970514 19981207	NO	1997-2213		19970514
NO 304381 PT 102022 CZ 289923 JP 11071395 JP 3207155	B B6 A2 B2	20001229 20020417 19990316 20010910	CZ	1997-102022 1997-2196 1998-103178		19970626 19970710 19980414
GR 3031361 GR 3032771 JP 2004049235 PRIORITY APPLN. INFO.:	T3 T3 A2	20000131 20000630 20040219	GR JP	1999-402455 2000-400473 2003-180211 1989-398667	A	19990929 20000228 20030624 19890825
·			US US US	1990-611419 1990-611965 1991-758880	A A A	19901108 19901108 19910913
			US US	1987-122714 1987-139886 1988-161072 1988-191263	B2 B2	19871118 19871230 19880226 19880506
			US US	1988-263584 1988-271450 1989-325338 1989-341334	B2 B2	19881026 19881114 19890317 19890420
			US US US	1989-353896 1989-355002 1989-355961	B2 B2 B2	19890421 19890518 19890518
			US JP	1989-456637 1990-504352 1990-512531	A A3	19891221 19900404 19900822
			WO WO	2001-75114 1990-US4766 1991-US2225 1991-302910	A A	19900822 19900822 19910329 19910403
			CZ EP	1991-2095521 1993-824 1992-900091	A3 A3	19911107 19911107 19911107
			JP JP	1997-120661 1992-500944 1998-103178 2001-59335	A3 A3	19911107 19911107 19911107 19911107
			US FI	1991-US8272 1992-910760 1993-2025 1993-97853	Α	19911107 19920707 19930505 19930727
7D			U.S	1773 77033	ΑI	1990121

AB Two hepatitis C virus (HCV) envelope proteins (E1 and E2) are manufactured without sialylation. Expression of these genes in lower eukaryotes, or in mammalian cells in which terminal glycosylation is blocked, results in proteins similar to native HCV glycoproteins. When isolated by mannose-binding GNA (Galanthus nivalus agglutinin) lectin affinity, the E1 and E2 proteins aggregate into virus-like particles. Cells bearing a mannose receptor or asialoglycoprotein receptor are capable of being infected with HCV and of supporting culturing of the virus. E1 and E2 were produced in HeLa S3 cells inoculated with recombinant Vaccinia virus containing HCV gene fragments and purified using a GNA-agarose column.

=> D L11 IBIB ABS 1-5

AUTHOR(S):

L11 ANSWER 1 OF 5 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 2002:180436 CAPLUS

DOCUMENT NUMBER: 137:227162

TITLE: Cloning and expression of human CD81 major

extracellular loop in E. coli and its activity Zhang, Guojun; Ling, Shigan; Song, Xiaoquo; Zhang,

Heqiu; Chen, Kun; Zhu, Cuixia; Xiu, Bingshui

CORPORATE SOURCE: Institute of Basic Medical Sciences, Academy of

Military Medical Sciences, Beijing, 100850, Peop. Rep.

China

SOURCE: Junshi Yixue Kexueyuan Yuankan (2001), 25(4), 260-264

CODEN: JYKYEL; ISSN: 1000-5501

PUBLISHER: Junshi Yixue Kexueyuan Yuankan Bianjibu

DOCUMENT TYPE: Journal LANGUAGE: Chinese

An expression plasmid for a fusion protein of human CD81 major extracellular loop was constructed and binding activity of its expressed protein with HCV E2 was studied. CD81 major extracellular loop sequence was amplified from human peripheral blood lymphocytes by RT-PCR, then inserted into the expression vector pBVIL1, and expressed in E. coli. The purified fusion protein was tested for binding activity with E2. CD81-EC2 gene was correctly amplified and inserted into the vector as confirmed by sequencing. The preliminary study showed that the recombinant CD81/EC2 could bind truncated

HCV E2 (384-661) protein expressed in E. coli. This work proved the way for further study on interactions of CD81 with HCV and its E2, and for preparation of anti-EC2 monoclonal antibody.

L11 ANSWER 2 OF 5 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 2001:912910 CAPLUS

DOCUMENT NUMBER: 137:104371

TITLE: Secretory expression of different C-terminal

truncated HCV El proteins in

mammalian cells and characterization of the expressed

products

AUTHOR(S): Zhu, Jun; Kong, Yuying; Liu, Jing; Zhang, Zuchuan;

Wang, Yuan; Li, Guangdi

CORPORATE SOURCE: Institute of Biochemistry and Cell Biology, Shanghai

Institute for Biological Sciences, Chinese Academy of

Sciences, Shanghai, 200031, Peop. Rep. China

SOURCE: Shengwu Huaxue Yu Shengwu Wuli Xuebao (2001), 33(6),

634-640

CODEN: SHWPAU; ISSN: 0582-9879 Shanghai Kexue Jishu Chubanshe

DOCUMENT TYPE: Journal LANGUAGE: Chinese

PUBLISHER:

Three fragments of HCV envelope 1 (E1) with different C-terminal truncation at aa310, aa325, aa340 were cloned into the mammalian expression vector pSecTagB. An epitope in the hepatitis B surface antigen, preS1(21-47), were genetically engineered onto the N-terminus of the recombinant protein and used as an affinity tag for detection and purification The resulting pSec-preS1-Elt310, pSec-preS1-E1t325, and pSec- preS1-E1t340 were transiently expressed in the HeLa cells and antigenicity, secretory efficiency, and glycosylation type of the recombinant El proteins were compared. All of the three recombinant proteins could be detected by both preS1 monoclonal antibody and El polyclonal antiserum. The expression products were secreted and highly mannose-type glycosylated, with S1Elt325 being secreted, indicating the influence of the hydrophobic regions on the secretion of the El protein. Three CHO cell lines expressing the proteins, S1E1t310, S1E1t325, and S1E1t340, were established and CHO/pSecS1E1t325 was chosen for further study. The secreted S1E1t325 could be enriched from cell culture medium by the preS1 antibody-coupled Sepharose. The glycosylation anal. indicated the lack of complex glycogen

even after the E1 was secreted via Golgi complexes. The established stable cell lines and anti-preS1 affinity method could be utilized to

may be used for further characterization of this protein.

L11 ANSWER 3 OF 5 CAPLUS COPYRIGHT 2005 ACS on STN

1998:592014 CAPLUS

enrich and purify the HCV El expressed in mammalian cells, and

DOCUMENT NUMBER: 129:301407

ACCESSION NUMBER:

TITLE: Hepatitis C virus envelope DNA-based

immunization elicits humoral and cellular immune

responses

AUTHOR(S): Lee, Seung Woo; Cho, Jae Ho; Lee, Ki Jeong; Sung,

Young Chul

Department of Life Science, Center for Biofunctional CORPORATE SOURCE:

Molecules, School of Environmental Engineering, Pohang University of Science and Technology, Pohang, 790-784,

S. Korea

Molecules and Cells (1998), 8(4), 444-451 SOURCE:

CODEN: MOCEEK; ISSN: 1016-8478

PUBLISHER: Springer-Verlag Singapore Pte. Ltd.

DOCUMENT TYPE: Journal English LANGUAGE:

The vaccine development for hepatitis C virus (HCV) is highly

urgent to prevent non A and non B hepatitis. It was recently shown that the HCV envelope proteins appeared to the key viral

antigens to induce protective immunity. To generate immune responses to

the HCV envelope proteins on the DNA-based

immunization, various envelope gene-containing plasmids were constructed. For efficient expression and secretion of envelope proteins, the signal sequence of each envelope protein was replaced with either herpes simplex virus type-1 (HSV-1) gD or signal sequence of gD and truncated C-terminal hydrophobic regions of envelope proteins. The i.m. injection of these plasmids generated a significant level of antibody titers to the E1 and E2 proteins, which maximally reached 850 and 25,000 resp. The secreted form of each envelope protein and the fusion of the highly immunogenic gDproteins were shown to have no significant effect on generating immune responses to the envelope proteins. In addition, immunized rats appeared to generate antibodies directed to the homologous HVR-1 peptide. Splenic lymphocytes from immunized rats were shown to induce significant T-cell proliferative responses with the stimulation of recombinant E1 and E2 proteins. Our results demonstrated that the HCV envelope-DNA based immunization could elicit both humoral and cellular immune responses.

REFERENCE COUNT: THERE ARE 46 CITED REFERENCES AVAILABLE FOR THIS 46 RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L11 ANSWER 4 OF 5 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 1997:113448 CAPLUS

DOCUMENT NUMBER:

126:117059

TITLE:

Method for detection of antibody to hepatitis C virus

second envelope glycoprotein

Okasinski, Gregory F.; Schaefer, Verlyn G.; Suhar, INVENTOR(S):

Thomas S.; Lesniewski, Richard R.; Scheffel, James W.

PATENT ASSIGNEE(S):

Abbott Laboratories, USA

SOURCE:

PCT Int. Appl., 34 pp.

CODEN: PIXXD2

DOCUMENT TYPE:

Patent

LANGUAGE:

English

FAMILY ACC. NUM. COUNT:

PATENT INFORMATION:

PATENT NO.	KIND DATE	APPLICATION NO.	DATE
WO 9641196	A1 19961219	WO 1996-US8536	19960604
· · · · · · · · · · · · · · · · · · ·		FR, GB, GR, IE, IT, I	
CA 2223277 EP 836708		CA 1996-2223277 EP 1996-917969	19960604 19960604
R: AT, BE, CH, JP 11507129	DE, ES, FR, GB, T2 19990622	IT, LI, NL JP 1996-501105	19960604
PRIORITY APPLN. INFO.:		US 1995-481018 WO 1996-US8536	A 19950607 W 19960604

AB A method for detecting antibody to HCV in a test sample. method includes utilizing a recombinant protein that is the expression product of mammalian cells transformed by a heterologous expression vector comprising a DNA sequencing encoding an E2 truncated protein. Test kits which include this recombinant protein also are provided.

ACCESSION NUMBER: 1996:698698 CAPLUS

DOCUMENT NUMBER: 126:6277

TITLE: Expression of HCV envelope

proteins and the serological utility of the anti-E2

immune response

AUTHOR(S): Lesniewski, Richard R.; Watanabe, Shinichi; Devare,

Sushil G.

CORPORATE SOURCE: Hepatitis Research and Development, Abbott

Laboratories, Abbott Park, IL, 60064, USA

Proceedings of the International Symposium of the SOURCE:

> Princess Takamatsu Cancer Research Fund (1995), Volume Date 1994, 25th (Hepatitis C Virus and Its Involvement

in the Development of Hepatocellular Carcinoma),

129-137

CODEN: PPTCBY

PUBLISHER: Princeton Scientific

DOCUMENT TYPE: Journal LANGUAGE: English

The 5' end of the hepatitis C virus (HCV) genome encodes

structural proteins of the virion. The first gene encodes a highly basic core protein. Immediately downstream of the core gene are regions which

encode the envelope proteins (E1 and E2) of the virus.

Artificial expression and secretion of immunol. active envelope proteins have proven to be a substantial challenge due to the high degree of glycosylation and the existence of certain hydrophobic domains contained within these sequences. Bacterial cell expression of

recombinant HCV envelope proteins results in

products that are not glycosylated and are poorly immunogenic. Emphasis has shifted to the use of mammalian cell lines (human embryonic kidney [HEK] and Chinese hamster ovary [CHO] cells) for the expression of glycosylated, immunol. active envelope proteins. Using HEK cells, E1 is expressed intracellularly but is not secreted from the cells. When El is cloned in fusion with a C-terminal truncated E2 protein, both proteins are detected intracellularly; however, only E2 is secreted. When the E1/E2 processing site is interrupted by constructing deletion mutants, the unprocessed E1/E2 fusion protein can be secreted from the cells. Quantifiable expression and secretion of a truncated E2 protein is now possible using CHO cells and SV40-based vectors. The HCV E2 glycoprotein expressed from CHO cells is highly antigenic; a strong humoral response to this

antigen develops in persons infected with HCV. Antibodies to E2 are found in 95% of patients with detectable HCV RNA in their sera. The presence of antibodies to E2 is not indicative of viral clearance and therefore the role these antibodies play in protective immunity, if any, is unclear.

=> D L9 IBIB ABS 1-9

ANSWER 1 OF 9 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER:

2004:402741 CAPLUS

DOCUMENT NUMBER:

140:373891

TITLE:

Recombinant hepatitis C virus El and E2 envelope proteins for diagnostic and

therapeutic use

INVENTOR(S):

Maertens, Geert; Bosman, Fons; Buyse, Marie Ange

PATENT ASSIGNEE(S): Belg.

SOURCE:

U.S. Pat. Appl. Publ., 162 pp., Cont.-in-part of U.S.

Ser. No. 355,040.

CODEN: USXXCO

DOCUMENT TYPE:

LANGUAGE:

Patent English

FAMILY ACC. NUM. COUNT:

PATENT INFORMATION:

PATENT NO.	KIND	DATE	APPLICATION NO.	DATE
US 2003118603 WO 9967285	A1 A1	20030626 19991229	US 2001-995860 WO 1999-EP4342	20011129 19990623

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AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CU, CZ,
             DE, DK, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS,
             JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MD, MG, MK,
             MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ,
             TM, TR, TT, UA, UG, US, UZ, VN, YU, ZA, ZW, AM, AZ, BY, KG, KZ,
             MD, RU, TJ, TM
         RW: GH, GM, KE, LS, MW, SD, SL, SZ, UG, ZW, AT, BE, CH, CY, DE, DK,
             ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, BF, BJ, CF, CG,
             CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG
     US 6635257
                                20031021
                                            US 1999-355040
                                                                   19990723
                          В1
                                20030310
                                            ZA 2000-7318
                                                                   20001208
     ZA 2000007318
                          Α
     TR 200202169
                          T1
                                20040621
                                            TR 2002-200202169
                                                                   20020111
                                20040213
                                            ZA 2002-7272
     ZA 2002007272
                         Α
                                                                   20020910
                                            EP 1998-870142
PRIORITY APPLN. INFO.:
                                                               A 19980624
                                            EP 1999-870033
                                                               A 19990222
                                            WO 1999-EP4342
                                                                W 19990623
                                            US 1999-355040
                                                               A2 19990723
                                            US 2000-304194P
                                                               P 20001201
                                                               P 20010111
                                            US 2001-260669P
                                            US 2001-315768P
                                                               P 20010830
     The present invention relates to a method for purifying
AB
     recombinant HCV single or specific oligomeric
     envelope proteins selected from the group consisting of El and/or
     E2 and/or E1/E2, characterized in that upon lysing the transformed host
     cells to isolate the recombinantly expressed protein a disulfide bond
     cleavage or reduction step is carried out with a disulfide bond cleavage
     agent. The present invention also relates to a composition isolated by such a
    method. The present invention also relates to the diagnostic and
     therapeutic application of these compns. Furthermore, the invention
     relates to the use of HCV El protein and peptides for prognosing
     and monitoring the clin. effectiveness and/or clin. outcome of HCV
     treatment.
    ANSWER 2 OF 9 CAPLUS COPYRIGHT 2005 ACS on STN
                         2003:756030 CAPLUS
ACCESSION NUMBER:
DOCUMENT NUMBER:
                         139:349007
                         Lethality in mice infected with recombinant
TITLE:
                         vaccinia virus expressing hepatitis C virus
                         core protein
AUTHOR(S):
                         Zhang, Hong
                         ISIS Pharmaceuticals, Carlsbad, CA, 92008, USA
CORPORATE SOURCE:
SOURCE:
                         Hepatobiliary & Pancreatic Diseases International
                         (2003), 2(3), 374-382
                         CODEN: HPDIAJ; ISSN: 1499-3872
PUBLISHER:
                         First Affiliated Hospital, Zhejiang University School
                         of Medicine
DOCUMENT TYPE:
                         Journal
LANGUAGE:
                         English
    OBJECTIVE: To establish a mouse model of HCV core expression and
     investigate the toxicity of HCV core protein or the possible
     pathogenic effects. METHODS: A series of vaccinia viral
     expression vectors were engineered to express 5' portion of
     HCV genes including 5' non-translated region (NTR), core protein,
     and portion of the El gene. These HCV sequences were fused to a
     luciferase reporter gene and inserted into a vaccinia virus
     expression vector (pSC11) adjacent to the vaccinia
     virus promoter, p7.5. The recombinant DNA constructs were
     packed into infectious recombinant chimeric viruses.
     expression of HCV core protein was examined in cultured cells
     after infection with these viruses. Death of the infected mice was
     investigated by specific correlation to the expression of HCV
     core protein and its expression levels. RESULTS: The recombinant
     virus (VNCE-LUA) expressed HCV core protein and an
     envelope-luciferase fusion protein in cultured cells. When Balb/c
    mice were inoculated i.p. with more than 107 pfu per mouse of VNCE-LUA,
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death occurred immediately. The mortality was dependent on the amount of

VNCE-LUA died within 4 days of infection and 50% of mice inoculated with 3

VNCE-LUA virus inoculated. All mice inoculated with 3 + 108 pfu of

+ 107 pfu of VNCE-LUA died within 7 days of infection. No death

occurred in mice inoculated with 3 + 108 pfu of a control recombinant vaccinia virus, which expressed luciferase but not the HCV core and envelope proteins. Deletion of core sequences from VNCE-LUA rapidly reduced the mortality of infected mice whereas deletion of envelope sequence did not. SCID mice infected with VNCE-LUA died 2-3 days after infection, suggesting that the HCV-core induced mortality is not dependent on host T- or B-cell responses to core protein. CONCLUSIONS: HCV core protein can be lethal to mice when expressed in vivo and this specific lethality is independent of T-cells or B-cells. The findings and model itself provide a useful tool for further investigation on potential pathol. effects as well as the potential toxicity of the HCV core protein.

REFERENCE COUNT: 29 THERE ARE 29 CITED REFERENCES AVAILABLE FOR THIS RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

L9 ANSWER 3 OF 9 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 2003:491258 CAPLUS

DOCUMENT NUMBER: 139:67765

TITLE: Recombinant hepatitis C virus E1 and E2

envelope proteins for diagnostic and

therapeutic use

INVENTOR(S): Maertens, Geert; Depla, Erik; Bosman, Fons

PATENT ASSIGNEE(S): Innogenetics N.V., Belg. SOURCE: PCT Int. Appl., 270 pp.

CODEN: PIXXD2

DOCUMENT TYPE: Patent LANGUAGE: English

FAMILY ACC. NUM. COUNT: 1

PATENT INFORMATION:

PA	PATENT NO.				KIND DATE					ICAT				DATE			
WO	2003	0519	12		A2 20030626 A3 20040304 C2 20040715									20021218			
	₩:	CO, GM, LS, PL,	CR, HR, LT, PT,	CU, HU, LU, RO,	CZ, ID, LV, RU,	DE, IL, MA, SC,	AU, DK, IN, MD, SD,	DM, IS, MG, SE,	DZ, JP, MK, SG,	EC, KE, MN, SK,	EE, KG, MW, SL,	ES, KP, MX,	FI, KR, MZ,	GB, KZ, NO,	GD, LC, NZ,	GE, LK, OM,	GH, LR, PH,
	RW:	GH, KG, FI,	GM, KZ, FR,	KE, MD, GB,	LS, RU, GR,	MW, TJ, IE,	VN, MZ, TM, IT, GN,	SD, AT, LU,	SL, BE, MC,	SZ, BG, NL,	TZ, CH, PT,	CY, SE,	CZ, SI,	DE, SK,	DK, TR,	EE,	ES,
	2468				AA		2003										
	2004 1461						2004									0021. 0021.	
BR NZ	R: 2002 5333 2005	AT, IE, 0150 96 5169	BE, SI, 81	CH, LT,	DE, LV, A A	DK, FI,	ES, RO, 2004	FR, MK, 1019 0429	GB, CY,	GR, AL, BR 2 NZ 2 JP 2 US 2 US 2	IT, TR, 002-1 002-1 003-1 001-1	LI, BG, 1508: 5333: 5527: 2051: 4183:	LU, CZ, 1 96 92 0	NL, EE,	SE, SK 2 2 2 2 A 2 P 2	MC, 0021: 0021:	PT, 218 218 218 218 218
_						_			_						_		-

The present invention relates to a method for purifying recombinant HCV single or specific oligomeric envelope proteins selected from the group consisting of E1 and/or E2 and/or E1/E2, characterized in that upon lysing the transformed host cells to isolate the recombinantly expressed protein a disulfide bond cleavage or reduction step is carried out with a disulfide bond cleavage agent. The present invention also relates to a composition isolated by such a method. The present invention also relates to the diagnostic and therapeutic application of these compns. Furthermore, the invention relates to the use of HCV E1 protein and peptides for prognosing and monitoring the clin. effectiveness and/or clin. outcome of HCV treatment.

ANSWER 4 OF 9 CAPLUS COPYRIGHT 2005 ACS on STN L9

2000:467550 CAPLUS ACCESSION NUMBER:

DOCUMENT NUMBER: 133:236484

Induction of hepatitis C virus-specific cytotoxic T TITLE:

lymphocytes in mice by an intrahepatic inoculation

with an expression plasmid

Kamei, Akira; Tamaki, Shigenori; Taniyama, Hiroyuki; AUTHOR(S):

Takamura, Shiki; Nishimura, Yuki; Kagawa, Yumiko; Uno-Furuta, Satori; Kaito, Masahiko; Kim, Gisen; Toda,

Masaaki; Matsuura, Yoshiharu; Miyamura, Tatsuo;

Adachi, Yukihiko; Yasutomi, Yasuhiro

Department of Bioregulation, Mie University School of CORPORATE SOURCE:

> Medicine, Mie, 514-8507, Japan Virology (2000), 273(1), 120-126

CODEN: VIRLAX; ISSN: 0042-6822

PUBLISHER: Academic Press

DOCUMENT TYPE: Journal LANGUAGE: English

SOURCE:

The authors assessed the possibility of intrahepatic inoculation with a plasmid encoding hepatitis C virus (HCV) proteins to elicit

HCV-specific cytotoxic T lymphocytes (CTL) in mice as a conventional animal model of HCV infection. BALB/c mice were intrahepatically or i.m. inoculated with an expression plasmid DNA encoding HCV structural proteins under the control of the elongation factor $1-\alpha$ promoter. Expressions of HCV-core protein and envelope proteins (E1 and E2) in hepatocytes were

detected immunohistochem. 6 days after inoculation. CTL responses were examined using target cells either pulsed with a specific peptide or

infected with a recombinant vaccinia virus expressing HCV structural protein. Both intrahepatically and i.m.

DNA-inoculated mice developed CD8+, MHC class I-restricted CTL responses

that recognized the peptide pulsed as well as HCV proteins

expressing target cells. These studies demonstrated the usefulness of a

murine model of HCV infection induced by direct intrahepatic DNA inoculation for understanding the immunopathogenic mechanisms in

HCV infection. (c) 2000 Academic Press.

THERE ARE 27 CITED REFERENCES AVAILABLE FOR THIS REFERENCE COUNT: 27

RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

ANSWER 5 OF 9 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 1998:205500 CAPLUS

128:290843 DOCUMENT NUMBER:

Expression of structural proteins of hepatitis C virus TITLE:

(HCV) in mammalian cells

Li, Yingchun; Li, Guangdi; Kong, Yuying; Wang, Yuan; AUTHOR(S):

Wang, Yu; Wen, Yumei

Shanghai Inst. Biochemistry, Chinese Academy Sciences, CORPORATE SOURCE:

Shanghai, 200031, Peop. Rep. China Science in China, Series C: Life Sciences (1998), SOURCE:

41(1), 47-55 CODEN: SCCLFO; ISSN: 1006-9305

Science in China Press PUBLISHER:

DOCUMENT TYPE: Journal LANGUAGE: English

The vaccinia viral vector containing T7 promoter was used

to construct the expression plasmids carrying HCV structural genes of C, El and E2/NS1. These genes were transiently expressed in

mammalian cells in the presence of the T7 RNA polymerase which was

provided by the recombinant vaccinia virus vTT7.

Expression of mature core protein, envelope protein E1 and E2 was detected by Western blot using HCV patient sera as the

primary antibodies. It was found that the sera from different HCV

patients reacted differently with the expressed products, so did the sera collected at different times from the same patient, from whom the HCV structural genes were isolated. Among six mammalian cell

lines, Vero and HeLa were the most suitable for the expression of C, E1

and E2. The recombinant vaccinia viruses have been constructed to constantly produce the C, E1 and E2 proteins for further

research.

L9 ANSWER 6 OF 9 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 1996:295079 CAPLUS

DOCUMENT NUMBER: 124:352673

Recombinant production and purification of TITLE: hepatitis C virus envelope proteins for

diagnostic and therapeutic use

INVENTOR(S): Maertens, Geert; Bosman, Fons; De Martynoff, Guy;

Buyse, Marie-Ange

Innogenetics N.V., Belg. PATENT ASSIGNEE(S): PCT Int. Appl., 146 pp. SOURCE:

CODEN: PIXXD2

DOCUMENT TYPE:

Patent

LANGUAGE:

English

FAMILY ACC. NUM. COUNT: PATENT INFORMATION:

PATENT NO.							DATE APPLICATION NO.					NO.	DATE			
	9604385			A2		1996	0215	1	WO 1	995-	EP30	31		:	19950	731
WO				A3		1996			011	017	0 B	D.D.	D.//		П.О	-
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	TT,		1-144	MA,	NO,	NZ,	FШ,	F1,	NO,	RU,	3U,	SE,	36,	31,	SK,	10,
	RW: KE,		SD.	S7.	HG	ΔТ.	BE	CH	DE.	DΚ	ES	FR	GB	GR	TE	TΨ
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CA	2172273	,		AA		1996	0215	1	CA 1	995-	2172	273			19950	731
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AU	708174			В2		1999	0729									
	721505			A1 B1		1996	0717		EP 1	995-	9304	34		:	19950	731
EP	721505					2002										
	R: AT,	BE,	CH,	DE,	DK,	ES,	FR,								NL,	
	09503396			T2		1997									19950	
	9506059			A		1997 2000	1028		BR 1	.995-	6059			:	L9950 L9950	731
	71728			A A1 E		2000	0418		SG 1	.997-	3877				19950	731
															9950	
EP	1211315		~	A1		2002				002-					19950	
D.M.	R: AT,	BE,	CH,													
	721505			T		2002	1031		PT 1	995-	9304	34			19950 19950	/31 721
	2174957 6150134			T3		2002			LO 1	995-	6120	34 72		:	19950 19960	731
	6245503			A. D1		2000		,	US 1	-066. -202	0129	/ 3 97			19900	011
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	757962			B2		2003			1 11 A	999-	5712	7			9970 9991	029
	9957127			A1		2000		•			J / 12	'		-		023
	20030361	10		A1		2003		1	US 2	001-	8993	03		2	20010	706
	20021827			A1		2002		1	US 2	001-	9730	25		- 2	20011	010
	200309598	80		A1		2003		1	US 2	001-	9958	08		2	20011 20011 20040	129
JP	20042227	29		A2		2004	0812	,	Ĵ₽ 2	004-	5170	9		2	20040	226
US	20041850	61		A1		2004	0923	1	JS 2	004-	8252	19		2	20040	416
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US 2001-315768P P 20010830 US 2001-973025 A2 20011010

Envelope proteins E1 and E2 of hepatitis C virus (HCV AΒ), their recombinant production and purification, their fragments and engineered derivs., their antigenic epitope peptides, their monoclonal antibodies, and their use for diagnostic and therapeutic means are provided. A method is described for purifying recombinant HCV single or specific oligomeric envelope proteins, characterized in that upon lysing the transformed host cells to isolate the recombinantly expressed protein a disulfide bond cleavage or reduction step is carried out with a disulfide bond cleavage agent (such as dithiothreitol and/or Empigen BB) and an SH group protecting agent (such as N-ethylmaleimide). Various forms of the E1 and E2 proteins are constructed by standard genetic techniques using vaccinia virus recombination vectors; such proteins are specific for various HCV genotypes, may delete the hydrophobic region from E1, or remove various glycosylation sites; they may also add factor Xa cleavage sites and His6 tags for improved purification Epitope (such as F, G, H, and I) peptides are used to generate monoclonal antibodies and to monitor disease progression in patients. Furthermore, the HCV El protein and peptides are used for prognosing and monitoring the clin. effectiveness and/or clin. outcome of HCV treatment.

L9 ANSWER 7 OF 9 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 1994:602124 CAPLUS

DOCUMENT NUMBER: 121:202124

TITLE: Formation and intracellular localization of hepatitis

C virus envelope glycoprotein complexes

expressed by recombinant vaccinia

and Sindbis viruses

AUTHOR(S): Dubuisson, Jean; Hsu, Henry H.; Cheung, Ramsey C.;

Greenberg, Harry B.; Russell, David G.; Rice, Charles

Μ.

CORPORATE SOURCE: Sch. Med., Washington Univ., St. Louis, MO,

63110-1093, USA

SOURCE: Journal of Virology (1994), 68(10), 6147-60

CODEN: JOVIAM; ISSN: 0022-538X

DOCUMENT TYPE: Journal LANGUAGE: English

Hepatitis C virus (HCV) encodes two putative virion glycoproteins (E1 and E2) which are released from the polyprotein by signal peptidase cleavage. In this report, the authors have characterized the complexes formed between E1 and E2 (called E1E2) for two different HCV strains (H and BK) and studied their intracellular localization. Vaccinia virus and Sindbis virus vectors were used to express the HCV structural proteins in three different cell lines (HepG2, BHK-21, and PK-15). The kinetics of association between E1 and E2, as studied by pulse-chase anal. and copptn. of E2 with an anti-E1 monoclonal antibody, indicated that formation of stable E1E2 complexes is slow.' The times required for half-maximal association between E1 and E2 were 60 to 85 min for the H strain and more than 165 min for the BK strain. In the presence of nonionic detergents, two forms of E1E2 complexes were detected. The predominant form was a heterodimer of El and E2 stabilized by noncovalent interactions. A minor fraction consisted of heterogeneous disulfide-linked aggregates, which most likely represent misfolded complexes. Posttranslational processing and localization of the HCV glycoproteins were examined by acquisition of endoglycosidase H resistance, subcellular fractionation, immunofluorescence, cell surface immunostaining, and immunoelectron microscopy. HCV glycoproteins containing complex N-linked glycans were not observed, and the proteins were not detected at the cell surface. Rather, the proteins localized predominantly to the endoplasmic reticular network, suggesting

ANSWER 8 OF 9 CAPLUS COPYRIGHT 2005 ACS on STN

ACCESSION NUMBER: 1994:4188 CAPLUS DOCUMENT NUMBER: 120:4188

TITLE: Characterization of hepatitis C virus envelope

that some mechanism exists for their retention in this compartment.

glycoprotein complexes expressed by

recombinant vaccinia viruses

Ralston, Robert; Thudium, Kent; Berger, Kim; Kuo, AUTHOR(S):

Carol; Gervase, Barbara; Hall, John; Selby, Mark; Kuo,

George; Houghton, Michael; Choo, Qui Lim Chiron Corp., Emeryville, CA, 94608, USA Journal of Virology (1993), 67(11), 6753-61

CODEN: JOVIAM; ISSN: 0022-538X

Journal DOCUMENT TYPE: LANGUAGE: English

CORPORATE SOURCE:

SOURCE:

The authors constructed recombinant vaccinia virus vectors for expression of the structural region of hepatitis C

virus (HCV). Infection of mammalian cells with a vector (vv/HCV1-906) encoding C-E1-E2-NS2 generated major protein species of 22 kDa (C), 33 to 35 kDa (E1), and 70 to 72 kDa (E2), as observed previously with other mammalian expression systems. The bulk of the E1 and E2 expressed by vv/HCV1-906 was integrated into endoplasmic reticulum membranes as core-glycosylated species, suggesting that these E1 and E2 species represent intracellular forms of the HCV envelope proteins. HCV E1 and E2 formed E1-E2 complexes

which were precipitated by either anti-E1 or anti-E2 serum and which sedimented at approx. 15 S on glycerol d. gradients. No evidence of intermol. disulfide bonding between El and E2 was detected. El and E2 were copurified to approx. 90% purity by mild detergent extraction, followed by chromatog. on Galanthus nivalus lectin-agarose and DEAE-Fractogel. Immunization of chimpanzees with purified E1-E2 generated high titers of anti-E1 and anti-E2 antibodies. Further studies demonstrated that purified E1-E2 complexes were recognized at high frequency by HCV + human sera and generated protective immunity in chimpanzees, suggesting that these purified HCV envelope proteins display native HCV epitopes.

ANSWER 9 OF 9 CAPLUS COPYRIGHT 2005 ACS on STN

1992:528131 CAPLUS ACCESSION NUMBER:

DOCUMENT NUMBER: 117:128131

TITLE: Hepatitis C virus asialoglycoproteins manufacture for

vaccines or immunoassay

INVENTOR(S): Ralston, Robert O.; Marcus, Frank; Thudium, Kent B.;

Gervase, Barbara A.; Hall, John A.

PATENT ASSIGNEE(S): Chiron Corp., USA

SOURCE: PCT Int. Appl., 28 pp.

CODEN: PIXXD2

DOCUMENT TYPE: Patent LANGUAGE: English

FAMILY ACC. NUM. COUNT:

PATENT INFORMATION:

PA:	PENT NO.		APPLICATION NO.	DATE
WO	9208734		WO 1991-US8272 PL, RO, SU	19911107
	414475		GB, GR, IT, LU, NL, SE EP 1990-309120	19900821
	R: AT, BE, CH,	DE, DK, ES, FR,	GB, GR, IT, LI, LU, NL,	
ES	2110411	T3 19980216	AT 1990-309120 ES 1990-309120	19900821
CA	2064705	AA 19910226	CA 1990-2064705	19900822
		C 19990406	WO 1990-US4766	10000022
	W: AU, CA, JP		WO 1990-054766	19900822
ΑU	9063449	A1 19910403	AU 1990-63449	19900822
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			JP 1990-512531	
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	W: AU, BB, BG,	BR, CA, FI, GB,	HU, JP, KP, KR, LK, MC,	MG, MW, NO,
	PL, RO, SD,	SU		
	RW: BF, BJ, CF,	CG, CM, GA, ML,	MR, SN, TD, TG	
ΑU	9176510	A1 19911030	AU 1991-76510	19910329

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ΑU	639560		B2	19930729					
GB	2257784		A1	19930120	GB	1992-20480		19910329	
BR	639560 2257784 9106309 62706 217025 05508219 2733138 109916 172133		Α	19930420	BR	1992-20480 1991-6309		19910329	
HU	62706		A2	19930528	нп	1992-3146		19910329	
нп	217025		B	19991129		1772 0110		23320023	
TD	05508219		ш.э	19931118	TD	1991-507636		10010320	
TD	2722120		D2	19980330	ŲΓ	1991-307030		19910329	
JP	2/33138		B2	19980330		1075 00010		1001000	
RO	109916		BI	19950728	RO	1975-92012		19910329	
PL	1/2133		B1	19970829	PL	1991-296329		19910329	
RU	172133 2130969 450931 450931		C1	19990527	RU	1991-5053084 1991-302910		19910329	
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EP						1995-114016			
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	R. AT BE	Сн	DE	DK EG EB	GB GI	R, IT, LI, LU, 1991-302910 1991-302910 1995-114016 1995-114016 1991-2095521 1991-90267	NT.	SF.	
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ΑU	9190267		A1	19920611	AU	1991-90267		19911107	
ΑU	668078		В2	19960426					
EP	556292		A 1	19930825	EP	1992-900091		19911107	
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$_{ m PL}$	175610		В1	19990129	PL	1991-300038		19911107	
ΑТ	188220		F.	20000115	አጥ	1002 000001		10011107	
FS	2139591		πз	20000116	FC	1992 900091		19911107	
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CA	112440		D1	20000220	CA	1993-020		19911107	
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02	289006		В6	20011017	CZ	1993-824		19911107	
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	9203839		A	19921119		1992-3839		19921001	
	310241		B1		NO	1992-3039		19921001	
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$_{ m LT}$	3808		В	19960325	\mathtt{LT}	1993-1747		19931230	
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US	3808 5968775		B A	19991019	US	1993-1747 1995-438435		19950510	
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Р	R	Τ	O	R	I	Т	'Y	AF	Ρ.	$_{\rm LN}$	ΙN	F	O	:

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US	1989-398667	A	19890825
US	1990-611419	Α	19901108
US	1990-611965	Α	19901108
US	1991-758880	Α	19910913
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US	1987-139886	B2	19871230
US	1988-161072	B2	19880226
US	1988-191263	B2	19880506
US	1988-263584	B2	19881026
US	1988-271450	B2	19881114
US	1989-325338	B2	19890317
US	1989-341334	B2	19890420
US	1989-353896	B2	19890421
US	1989-355002	B2	19890518
US	1989-355961	B2	19890518
US	1989-456637	B2	19891221
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JΡ	1990-512531	A3	19900822
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EΡ	1991-302910	A3	19910403
CA	1991-2095521	A3	19911107
CZ	1993-824	A3	19911107
EΡ	1992-900091	A3	19911107
EΡ	1997-120661	A3	19911107
JΡ	1992-500944	A3	19911107
JΡ	1998-103178	A3	19911107
JP	2001-59335	A3	19911107
WO	1991-US8272	Α	19911107
US ·	1992-910760	A3	19920707
FI	1993-2025	Α	19930505
US	1993-97853	A1	19930727

AB Two hepatitis C virus (HCV) envelope proteins (El and E2) are manufactured without sialylation. Expression of these genes in lower eukaryotes, or in mammalian cells in which terminal glycosylation is blocked, results in proteins similar to native HCV glycoproteins. When isolated by mannose-binding GNA (Galanthus nivalus agglutinin) lectin affinity, the E1 and E2 proteins aggregate into virus-like particles. Cells bearing a mannose receptor or asialoglycoprotein receptor are capable of being infected with HCV and of supporting culturing of the virus. E1 and E2 were produced in HeLa S3 cells inoculated with recombinant Vaccinia virus containing HCV gene fragments and purified using a GNA-agarose column.